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Cell Culture Media Exosome Purification and RNA Isolation Midi Kit
Product Insert

Product # 60800

Exosomes are 40 - 150 nm membrane vesicles which are secreted by most cell types. Exosomes can be found in cell culture media, plasma, serum, saliva, urine, amniotic fluid and malignant ascite fluids, among other biological fluids. Evidence has been accumulating recently that these vesicles act as cellular messengers, conveying information to distant cells and tissues within the body. The exosomes contain cell-specific proteins, lipids and RNAs, which are transported to other cells, where they can alter function and/or physiology. These exosomes may play a functional role in mediating adaptive immune responses to infectious agents and tumours, tissue repair, neural communication and transfer of pathogenic proteins. Recent work has demonstrated the presence of distinct subsets of microRNAs within exosomes and other extracellular vesicles (EVs) which depend upon the tumour cell type from which they are secreted. For this reason exosomal RNA may serve as biomarkers for various diseases including cancer. Another subset of RNA that is found in cell culture media is the free-circulating RNA (fc-RNA). These fc-RNA are usually protein-bound RNA that are leaked from cells either during apoptosis or necrosis. As the RNA molecules encapsulated within exosomes or bound to proteins (fc-RNA) are protected from degradation by RNAses, they can be efficiently recovered from cell culture media. In general, these two RNA groups contain valuable information for the discovery of biomarkers that can help with early detection of certain cancer types and for monitoring the disease status.

Norgen's Cell Culture Media Exosome Purification and RNA Isolation Midi Kit constitutes an all-in-one system for the purification of exosomes and the subsequent isolation of exosomal RNA from different cell culture media sample volumes ranging from 10 mL to 20 mL. The purification is based on spin column chromatography that employs Norgen's proprietary resin. The kit is designed to isolate all sizes of extracellular vesicle RNA, including microRNA. The kit provides a clear advantage over other available kits in that it does not require any special instrumentation, protein precipitation reagents, extension tubes, phenol/chloroform or protease treatments. Moreover, the kit allows the user to elute into a flexible elution volume ranging from 50 µL to 100 µL. The purified RNA is free from any protein-bound circulating RNA and of the highest integrity. The purified RNA can be used in a number of downstream applications including real time PCR, reverse transcription PCR, Northern blotting, RNase protection and primer extension, and expression array assays.

Component	Product # 60800 (25 preps)
Slurry E	14.5 mL
ExoC Buffer	1.5 mL
ExoR Buffer	12 mL
Lysis Buffer A	20 mL
Lysis Additive B	2 mL
Wash Solution A	18 mL
Elution Solution A	6 mL
Mini Filter Spin Column	25
Mini Spin Columns	25
Collection Tubes	25
Elution tubes (1.7 mL)	50
Product Insert	1

¥ ***Please check page 4 for Average Yields and Common RNA Quantification Methods***

Storage Conditions and Product Stability

All buffers should be kept tightly sealed and stored at room temperature (15-25°C) for up to 2 year without showing any reduction in performance. It is recommended to warm Lysis Buffer A for 20 minutes at 60ºC if any salt precipitation is observed.

Customer-Supplied Reagents and Equipment

- Disposable powder-free gloves
- Benchtop microcentrifuge
- Micropipettors
- Sterile pipette tips with filters
- Vortex
- 96-100% Ethanol
- Nuclease-Free Water
- 15 mL tube

General Precautions

Proper biosafety measures should therefore be carried out when using this kit.

Quality Control

In accordance with Norgen's ISO 9001 and ISO 13485-certified Quality Management System, each lot of Norgen's Cell Culture Media Exosome Purification and RNA Isolation Midi Kit is tested against predetermined specifications to ensure consistent product quality.

Product Use Limitations

Norgen's Cell Culture Media Exosome Purification and RNA Isolation Midi Kit is designed for research purposes only. It is not intended for human or diagnostic use.

Product Warranty and Satisfaction Guarantee

NORGEN BIOTEK CORPORATION guarantees the performance of all products in the manner described in our product manual. The customer must determine the suitability of the product for its particular use.

Safety Information

Ensure that a suitable lab coat, disposable gloves and protective goggles are worn when working with chemicals. For more information, please consult the appropriate Material Safety Data Sheets (MSDSs). These are available as convenient PDF files online at *www.norgenbiotek.com*.

Lysis Buffer A contains guanidine thiocyanate, and should be handled with care. Guanidine thiocyanate forms highly reactive compounds when combined with bleach, thus care must be taken to properly dispose of any of this solution.

If liquid containing these buffers is spilled, clean with suitable laboratory detergent and water. If the spilled liquid contains potentially infectious agents, clean the affected area first with laboratory detergent and water, and then with 1% (v/v) sodium hypochlorite.

CAUTION: DO NOT add bleach or acidic solutions directly to the sample-preparation waste.

Important Notes

Working with RNA

RNases are very stable and robust enzymes that degrade RNA. Autoclaving solutions and glassware is not always sufficient to actively remove these enzymes. The first step when preparing to work with RNA is to create an RNase-free environment. The following precautions are recommended as your best defence against these enzymes.

- The RNA area should be located away from microbiological work stations
- Clean, disposable gloves should be worn at all times when handling reagents, samples, pipettes, disposable tubes, etc. It is recommended that gloves are changed frequently to avoid contamination
- There should be designated solutions, tips, tubes, lab coats, pipettes, etc. for RNA only
- All RNA solutions should be prepared using at least 0.05% DEPC-treated autoclaved water or molecular biology grade nuclease-free water
- Clean all surfaces with commercially available RNase decontamination solutions
- When working with purified RNA, ensure that they remain on ice during downstream applications

Notes Prior to Use

- All centrifugation steps are performed at room temperature.
- Ensure that centrifuge tubes used are capable of withstanding the centrifugal forces required.
- The provided spin columns are optimized to be used with a benchtop centrifuge and not to be used on a vacuum apparatus
- Most standard benchtop microcentrifuges will accommodate Norgen's Mini Spin Columns.
- Centrifuging Norgen's Spin Columns at a speed higher than recommended may affect RNA yield.
- Centrifuging Norgen's Spin Columns at a speed lower than recommended will not affect RNA yield. However, centrifugation at a lower speed may require longer time for the solutions to pass through the spin column
- Ensure that all solutions are at room temperature prior to use.
- It is highly recommended to warm up **Lysis Buffer A** at 60ºC for 20 minutes and mix well until the solutions become clear again if precipitates are present.
- Prepare a working concentration of the **Wash Solution A** by adding **42 mL** of 96 - 100% ethanol (provided by the user) to the supplied bottle containing **18 mL** of concentrated Wash Solution A. This will give a final volume of **60 mL.** The label on the bottle has a box that may be checked to indicate that the ethanol has been added.
- ***If any of the solutions do not go through the Spin Columns within the specified centrifugation time, spin for an additional 1-2 minutes until the solution completely passes through the Column. Do NOT exceed the centrifugation speed as this may affect RNA yield.***

Preparation of Cell-free Cell Culture Media Sample

1. Harvest and transfer the required cell culture media volume into a conical tube and centrifuge at **200 *x g* (~1,000 RPM) for 15 minutes** to remove any cells and debris.
2. Transfer cell-free media into a fresh 15-50 mL conical tube.

- **Cell-Free Cell Culture Media is now ready for Exosome purification**.

Section 1: Exosome Purification from 10 mL - 20 mL of Cell Culture Media

Note: The procedure outlined below is for 20 mL inputs of cell-free media. If processing a sample volume in the range of 10 mL - 20 mL media, simply add 2.5 µL ExoC Buffer for every 1mL of cell-free media. The volume of Slurry E and ExoR Buffer is constant for any volume to be processed.

1. To 20 mL cell-free media add 50 µL of **ExoC Buffer** followed by the addition of 400 µL of **Slurry E**. ***(Note: Mix Slurry E well prior to use. For optimal performance ensure that resin is completely resuspended).***
2. Mix well by vortexing for 10 seconds and let stand at room temperature for 10 minutes.
3. Mix well by vortexing for 10 seconds. Centrifuge for **2 minutes at 2,000 RPM**. Discard the supernatant.
4. Apply 400 µL **ExoR Buffer** to the slurry pellet and mix well by vortexing for 10 seconds.
5. Incubate the slurry pellet resuspended in the 400 µL **ExoR Buffer** at room temperature for 10 minutes.
6. After incubation, mix well by vortexing for 10 seconds then centrifuge for **2 minutes at 500 RPM**.
7. Transfer the supernatant to a Mini Filter Spin column assembled with an elution tube and centrifuge for 1 minute at 6,000 RPM. **Do Not Discard the flowthrough which contains your purified Exosomes.**
   - **Your Exosomes are now ready for any downstream applications.**

Section 2: Exosomal RNA Isolation

1. Add 600 µL of **Lysis Buffer A** and 75 µL of **Lysis Additive B** to the 400 µL ExoR Buffer containing the purified Exosomes (**Step 7, Section 1**).
2. Mix well by vortexing for 10 seconds then incubate at room temperature for 15 minutes
3. After Incubation add 1 mL of **96-100% Ethanol** to the mixture from **Step 2** and mix well by vortexing for 10 seconds.
4. Transfer 750 µL of the mixture from **Step 3** into a **Mini Spin Column**. Centrifuge for **1 minute at 3,300 *x g* (~6,000 RPM)**. Discard the flowthrough and reassemble the spin column with its collection tube.
5. Repeat **Step 4** two more times to transfer the remaining mixture from **Step 3** into the **Mini Spin Column.**

6. Apply 600 µL of **Wash Solution A** to the column and centrifuge for **30 seconds at 3,300 *x g* (~6,000 RPM)**. Discard the flowthrough and reassemble the spin column with its collection tube.
7. Repeat **Step 6** one more time, for a total of two washes.
8. Spin the column, empty, for **1 minute at 13,000 *x g* (~14,000 RPM)**. Discard the collection tube.
9. Transfer the spin column to a fresh 1.7 mL Elution tube. Apply 50 µL of **Elution Solution A** to the column and centrifuge for **1 minute at 2,000 RPM,** followed by **2 minutes at 8,000 RPM**.
10. For maximum recovery, transfer the eluted buffer back to the column and let stand at room temperature for 2 minutes. Centrifuge for **1 minute at 400 x *g* (~2,000 RPM),** followed by **2 minutes at 5,800 x g (~8,000 RPM)**.

    - **Exosomal RNA is now ready for downstream applications.**

Appendix A

Exosomal RNA Yield

Exosomal RNA from conditioned cell culture media is normally found in very low amounts (1 - 100 pg/µL), therefore measuring exosomal RNA concentration using common quantification methods is very difficult and challenging. Typical yields of exosomal RNA vary significantly depending on the number of exosomes that are shed from the cultured cells, as well as the kind of treatment the cultured cells are receiving, and therefore there is no absolute yield for RNA purified from conditioned cell culture media.

Below is a list of the most common RNA quantification methods, as well as the limit of detection for each of these methods. ***Unfortunately, none of these methods can be used reliably for measuring the concentration of RNA purified from exosomes unless large volumes have been processed***. This would only be applicable if exosomes contain the maximum amount of RNA that can fit within the specification range of these quantification tools. It should be noted that the specifications outlined below are based on measuring a pure RNA sample, which will not be the case for the exosomal RNA purified from media. Exosomal RNA is short fragmented RNA which is usually present in less than 1000 bp. Purified exosomal RNA usually contains traces of proteins which will interfere with most quantification methods, leading to the overestimation of the purified RNA concentration. Therefore purified RNA contaminated with more proteins will be presented at a higher concentration as compared to RNA purified with less protein contaminants, which in this case will depend on the method used for media RNA purification. ***The only reliable method that can assess the quality and the relative quantity of the purified plasma/serum RNA is RT-qPCR amplification of a standard RNA using a small RNA amplicon such as the 5S rRNA housekeeping gene***.

Common RNA Quantification Methods

1) Bioanalyzer RNA Quantification kits

	RNA 6000 Nano Kit		RNA 6000 Pico Kit		Small RNA kit
	Total RNA	**mRNA**	**Total RNA**	**mRNA**	**Total RNA**
Quantitative range	25 - 500 ng/µL	25 - 250 ng/µL	----	----	50-2000 pg/µL
Qualititative range	5 - 500 ng/µL	5 - 250 ng/µL	50 - 5000 pg/µL	250 - 5000 pg/µL	50-2000 pg/µL
Quantitation accuracy	20% CV	20% CV	30% CV	----	------

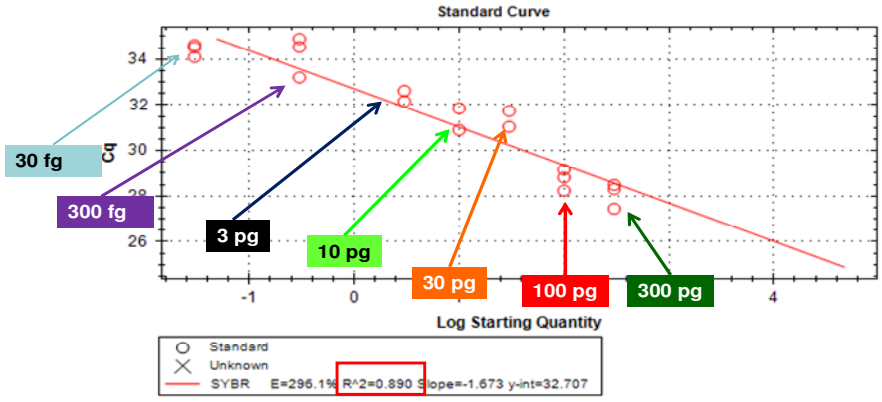
2) NanoDrop 2000

- Detection Limit: 2 ng/µl (dsDNA)

3) Quant-iT™ RiboGreen® RNA Assay Kit

- Quantitation Range: 1-200 ng

4) qPCR Standard Curve (generated by Norgen)



Frequently Asked Questions

1. **What If a variable speed centrifuge is not available?**
   - A fixed speed centrifuge can be used, however reduced yields may be observed.
2. **At what temperature should I centrifuge my samples?**
   - All centrifugation steps are performed at room temperature. Centrifugation at 4°C will not adversely affect kit performance.
3. **What if I added more or less of the specified reagents' volume?**
   - Adding more or less than the specified volumes may reduce both the quality and the quantity of the purified RNA. Eluting your RNA in high volumes will increase the yield but will lower the concentration. Eluting in small volumes will increase the concentration but will lower the overall yield.
4. **What if I forgot to do a dry spin before my final elution step?**
   - Your purified RNA will be contaminated with the Wash Solution A. This may reduce the quality of your purified RNA and will interfere with your downstream applications.
5. **Can I perform a second elution?**
   - Yes, but it is recommended that the 2nd elution be in a smaller volume (50% of 1st Elution). It is also recommended to perform the 2nd elution into a separate elution tube to avoid diluting the 1st elution.
6. **Why do my samples show low RNA yield?**
   - Exosomes contain very little RNA. This varies significantly depending on the number of exosomes that are shed from the cultured cells, as well as the kind of treatment the cultured cells are receiving. In order to increase the yield, the amount of media could be increased.
7. **Why is the A260/280 ratio of the purified RNA lower than 2.0?**
   - Most of the Exosomal RNA is short RNA fragments with a very low concentration where the A260/280 ratio tends to decrease with the decrease in the RNA concentration. The A260/280 ratio is normally between 1 – 1.6. This low A260/280 ratio will not affect any downstream application.
8. **Why does my isolated RNA not perform well in downstream applications?**
   - If a different Elution Buffer was used other than the one provided in the kit, the buffer should be checked for any components that may interfere with the application. Common components that are known to interfere are high salts (including EDTA), detergents and other denaturants. Check the compatibility of your elution buffer with the intended use.
9. **What should I do if some of the grey resin is transferred out when I am decanting the media supernatant?**
   - Simply remix and recentrifuge. After centrifuging decant the supernatant.
10. **What if I added more or less of Slurry E?**
    - Adding less volume may reduce the amount of the purified exosomes. Adding more may not affect the exosome capture but ma affect the release of the purified exosomes in the ExoR Buffer

11. What if I added more or less of ExoC Buffer?

- Adding a different volume from the specified optimum volume will signficiantly reduce the amount of the purified exosomes

12. What if I added more or less of ExoR Buffer?

- Adding less volume will reduce the release of the captured exosomes in the ExoR Buffer. Adding more will not affect the release of the captured exosomes but it will be more diluted

13. What will happen if some of the grey resin was accidently transferred with the ExoR buffer?

- Any grey resin will be filtered through the Midi Filter Spin Column and the flowthrough which contains the purified exosomes should not contain any grey resin.

Technical Assistance

NORGEN's Technical Service Department is staffed by experienced scientists with extensive practical and theoretical expertise in sample and assay technologies and the use of NORGEN products. If you have any questions or experience any difficulties regarding Norgen's Cell Culture Media Exosome Purification and RNA Isolation Midi Kit or NORGEN products in general, please do not hesitate to contact us.

NORGEN customers are a valuable source of information regarding advanced or specialized uses of our products. This information is helpful to other scientists as well as to the researchers at NORGEN. We therefore encourage you to contact us if you have any suggestions about product performance or new applications and techniques.

For technical assistance and more information, please contact our Technical Support Team between the hours of 8:30 and 5:30 (Eastern Standard Time) at (905) 227-8848 or Toll Free at 1-866-667-4362. or call one of the NORGEN local distributors (www.norgenbiotek.com) or through email at techsupport@norgenbiotek.com.

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